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Cord blood antibodies following maternal coronavirus disease 2019 vaccination during pregnancy

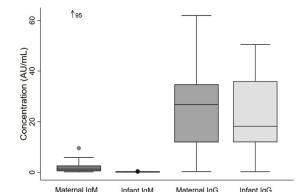
Vaccination of pregnant women can be an important strategy to confer protection to neonates and young infants. However, there are limited data on the immunologic response of pregnant women to the messenger RNA (mRNA) coronavirus disease 2019 (COVID-19) vaccinations² and the kinetics of transplacental

antibody transfer.^{3,4} Our objective was to investigate the transfer of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) immunoglobulin G (IgG) to infants maternal COVID-19 vaccination pregnancy and the factors associated with an increased efficiency of transfer.

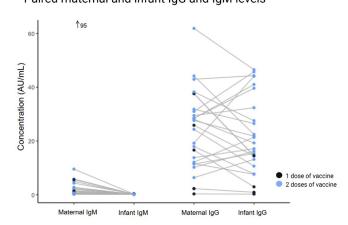
FIGURE

Maternal and infant SARS-CoV-2 antibodies and IgG transfer

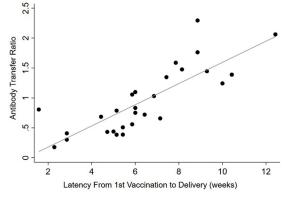
Maternal and infant SARS-CoV-2 antibody levels



В Paired maternal and infant IgG and IgM levels



Scatterplot of antibody transfer ratio and latency from vaccination to delivery



A, Anti—SARS-CoV-2 antibody levels in maternal and umbilical cord (infant) blood following maternal COVID-19 vaccination (n=27 mothers, n=28 infants); B, Paired maternal and infant IgM and IgG levels of women with 1 dose (n=5) and 2 doses (n=22) of the vaccine during pregnancy. Paired infant IgM was 0.2 for 1 outlier maternal IgM of 95 AU/mL; C, The association between the latency (weeks) from first vaccine dose and the antibody transfer ratio (infant IgG to maternal IgG).

COVID-19, coronavirus disease 2019; Ig, immunoglobulin; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2. Mithal. Antibody transfer after maternal coronavirus disease 2019 vaccination. Am J Obstet Gynecol 2021.

Research Letter

STUDY DESIGN: This was a prospective case series of pregnant women who delivered at the Prentice Women's Hospital in Chicago, IL (between January 2021 and March 2021). Women who received a COVID-19 vaccination during pregnancy were identified via review of the electronic health records (EHRs). Demographic and clinical information, including the specific vaccine used and the latency from vaccination to delivery was obtained from the EHRs. At the time of the study, 2 mRNA COVID-19 vaccines, the Pfizer (New York, NY) and Moderna (Cambridge, MA) vaccines, had received emergency use authorization, and healthcare workers were the initial qualifying group. Maternal blood and umbilical cord blood (herein referred to as "infant") were retrieved from specimens submitted for blood typing. SARS-CoV-2 IgM and IgG antibody levels were measured from the plasma using the Access SARS-CoV-2 IgG and IgM Antibody tests (DXI Platform, Beckman Coulter, Brea, CA) in a Certified Analytics Professional and Clinical Laboratory Improvement Amendments certified clinical laboratory. chemiluminescent assay quantitatively reports antibodies against the SARS-CoV-2 receptor binding domain in arbitrary units [AU]/mL, with values of ≥ 1 considered positive. The antibody transfer ratio was calculated as the infant IgG concentration divided by the maternal IgG concentration. Descriptive statistics were calculated. Linear regressions were used to identify factors significantly associated with the antibody transfer ratios and the infant IgG values. This study was institutional review board approved before its initiation.

RESULTS: We collected matched maternal plasma and cord blood from 27 women who received a COVID-19 vaccination during pregnancy; they delivered 28 infants (1 twin pair). The average maternal age was 33±3 (mean±standard deviation) years; race and ethnicity was 75% non-Hispanic white and 11% Hispanic. The average gestational age at first vaccine dose was 33 ± 2 weeks. For the vaccine type, 18 (64%) received the Pfizer vaccine, 6 (18%) received the Moderna vaccine, and 4 (14%) received a vaccine with the manufacturer unknown. Of note, 22 women (74%) received both vaccine doses before delivery with a mean latency of 6±3 weeks. About half of the women (15/27) and none of the infants had a positive IgM test (>1) (Figure, A). All but 1 woman (26/27) had a positive SARS-CoV-2 IgG test at the time of delivery. Only 3 infants did not have positive IgG tests (1 set of twins); these 2 women had received their first vaccine dose less than 3 weeks before delivery. The average maternal to infant IgG transfer ratio was 1.0±0.6 (Figure, B), however, an increased latency from vaccination to delivery (weeks) was associated with an increased transfer ratio $(\beta=0.2; 95\% \text{ confidence interval [CI]}, 0.1-0.2)$ (Figure, C). Correspondingly, having received the second vaccine dose before delivery was significantly associated with increased infant IgG levels (β =19.0; 95% CI, 7.1–30.8). Similarly, latency from vaccination to delivery was associated with increased infant IgG levels (β =2.9; 95% CI, 0.7-5.1).

CONCLUSION: Most pregnant women who received a COVID-19 mRNA vaccine during the third trimester had transplacental transfer of IgG to the infant. The observed mean IgG transfer ratio demonstrated that infant antibody levels are about equal to the maternal levels. This is slightly lower than what has been reported for the pertussis vaccinations (1.19-1.36),5 but greater than the transfer ratio following SARS-CoV-2 infection (0.72±0.57). A novel finding is that the transfer ratio seems to increase with latency from vaccination. These data suggest, at least among women in their third trimester, that earlier vaccination may produce a greater infant immunity, the immunobiology of which requires further study. Notably, owing to vaccine eligibility and timing, most women included herein were healthcare workers in their third trimester. Future research, including a more diverse cohort of women and women who received vaccination earlier in their pregnancy, is needed. Nevertheless, these results show promising evidence for passive immunity against SARS-CoV-2 in newborns after maternal receipt of COVID-19 mRNA vaccinations.

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The authors report no conflict of interest.

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